

## The Problem

Current Procedures are Not Designed for Limiting Amounts of DNA

Significant losses (>75%) in collection & purification steps

Assaying only ~5% of the PCR reaction products

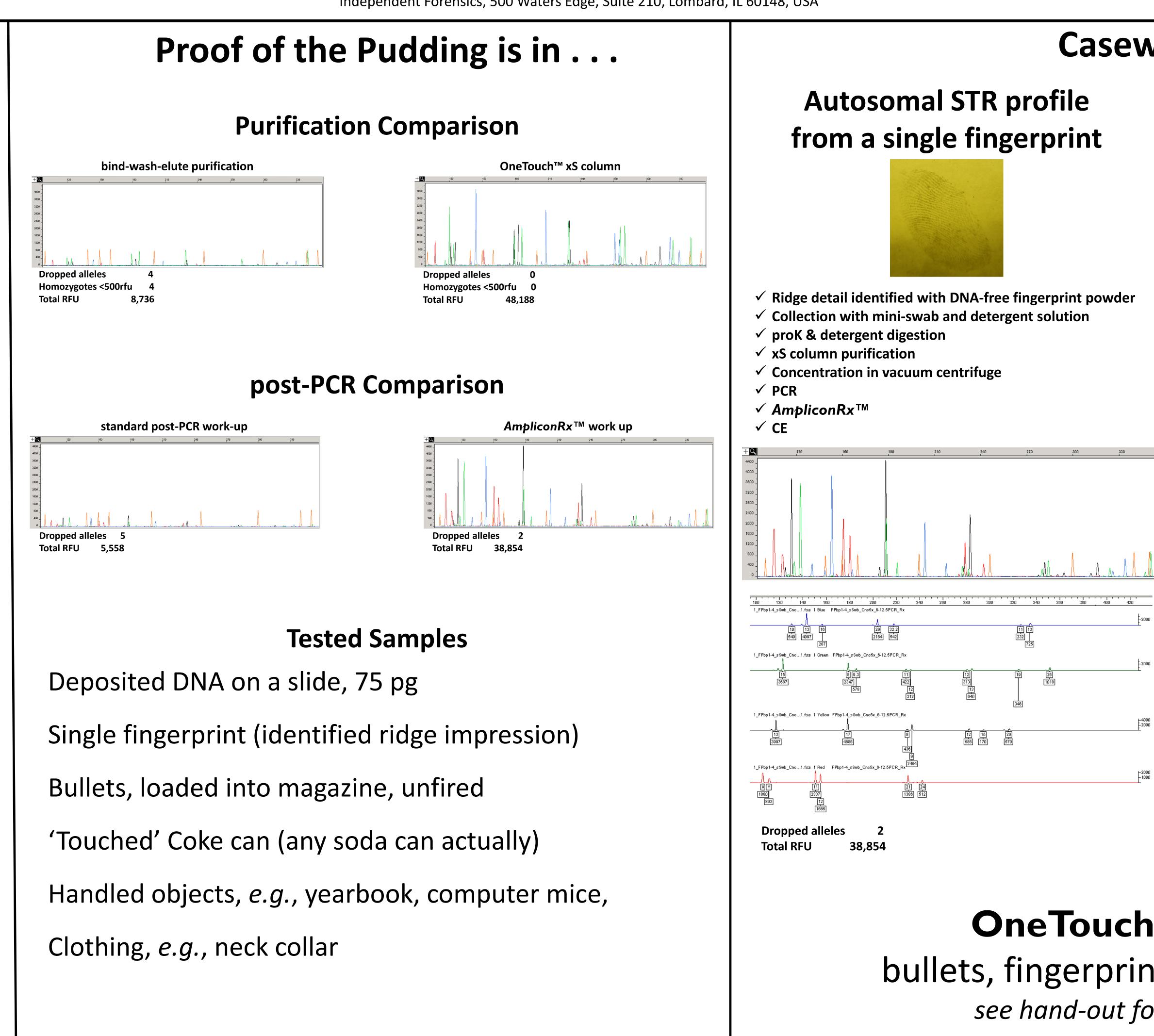
# The Solution

- **a**) collecting the biological material: mini-swabs and detergent-based buffer\* *\*secret sauce#1: better swabs and detergent*
- **b**) recovery of collected material: centrifugation at high speed
- c) release of DNA: proK & detergent at elevated °C
- **d**) purification of DNA: removal of inhibitors with 80-90% recovery\* \*secret sauce#2: NOT bind-wash-elute
- e) PCR amplification: modest changes, 29 v. 28 cycles, 2x TAQ\* \*better amp with no PCR drop-ins
- **f**) assay 100% of the PCR reaction: post-PCR processing and purification\* \*secret sauce#3: use all of the PCR reaction, Amplicon Rx<sup>™</sup>

## $\Sigma$ : the most sensitive technique, ever!

## **Obtaining DNA-Short Tandem Repeat (STR) Profiles from Evidentiary Samples with Extremely Limited Amounts of DNA**

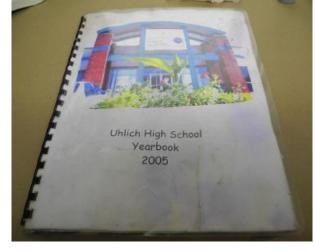
Alex Sinelnikov (alex@ifi-test.com), P.W. Boonlayangoor (boon@ifi-test.com) and Karl A. Reich (karl@ifi-test.com) Independent Forensics, 500 Waters Edge, Suite 210, Lombard, IL 60148, USA



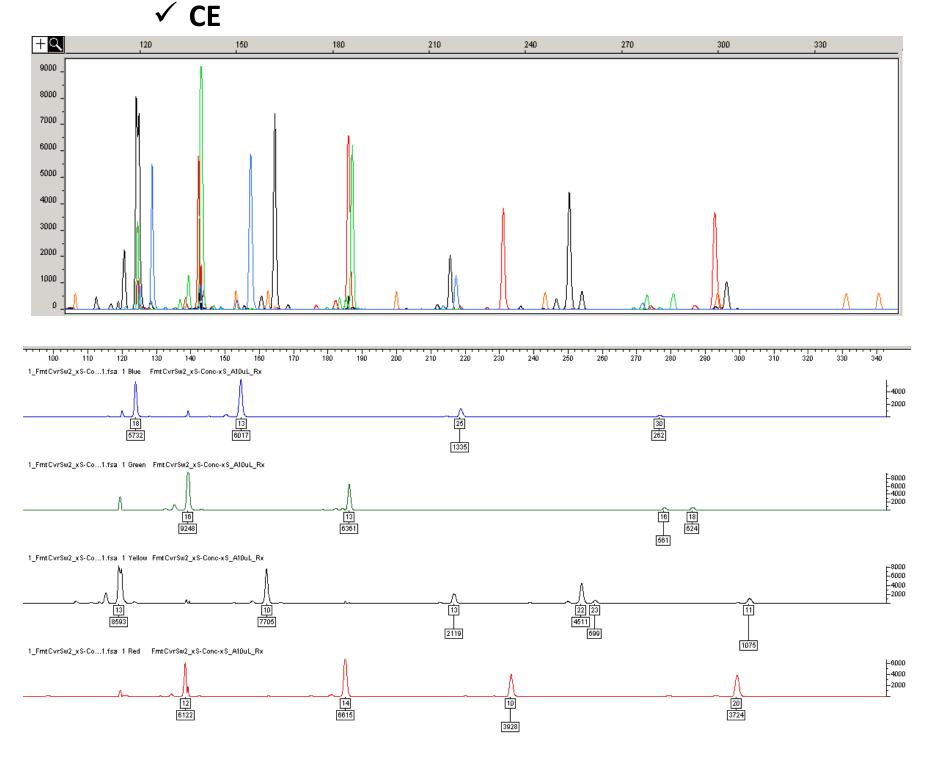
Poster #B9

#### **Casework Examples**

#### **Y** STR profile from a 'yearbook' cover



- ✓ Entire cover sampled with full-size swab & detergent solution (wet/dry technique)
- ✓ proK & detergent digestion
- $\checkmark$  xS column purification [1<sup>st</sup>]
- ✓ Concentration in vacuum centrifuge
- $\checkmark$  xS column purification [2<sup>nd</sup>]
- ✓ PCR
- ✓ AmpliconRx<sup>™</sup>



Dropped alleles Total RFU 77,532

#### **OneTouch DNA Method** bullets, fingerprints, handled objects... see hand-out for additional examples